# THE ROLE OF VITAMIN E (α-TOCOPHEROL) ON TESTOSTERONE LEVEL IN SPRAGUE DAWLEY RATS FOLLOWING CISPLATIN TREATMENT

<sup>1</sup>Dian Kartika Rezia, <sup>1</sup>Lukman Hakim, <sup>1</sup>Wahjoe Djatisoesanto.

<sup>1</sup>Department of Urology, Faculty of Medicine/Universitas Airlangga, Soetomo General Hospital, Surabaya.

#### **ABSTRACT**

**Objective:** To observe the difference of testosterone levels in adult male Sprague Dawley rats treated with combination of cisplatin and vitamin E compared to those treated with cisplatin only. **Material & Methods:** We used 24 adult male Sprague Dawley rats weight 200–300 grams and randomly assigned into 4 groups (n=6). Rats in negative control group (NC) were given intraperitoneal normal saline injection, while the positive control (PC) group were injected with cisplatin 5 mg/kgBW at the end of the 3<sup>rd</sup> week. Two other groups, P1 and P2, were injected with cisplatin 5 mg/kgBW and given vitamin E orally 50 mg/kgBW and 200 mg/kgBW, respectively. Cardiac blood was aspirated at the end of the 7<sup>th</sup> week and processed for analysis of testosterone levels. **Results:** Werecorded a significantly lower testosterone levels in rats treated only with cisplatin 5 mg/kgBW (PC) compared to those in CN group (P0.006), and those receiving combination of cisplatin and vitamin E 50 mg/kgBW (P0.003) and 200 mg/kgBW (P0.001). Though not significant, testosterone levels were higher in P2 group than in P1 group (P0.003). **Conclusion:** Exposure to cisplatin can lower testosterone levels in white rats, and the administration of vitamin E gives protection against such effect.

**Keywords:** Cisplatin, chemotherapy, vitamin E, antioxidant, testosterone, testis.

## **ABSTRAK**

Tujuan: Untuk mengetahui perbedaan kadar testosterone pada tikus Sprague Dawley yang diberikan kombinasi obat cisplatin dan vitamin E dibandingkan dengan tikus Sprague Dawley yang hanya mendapatkan cisplatin. Bahan & Cara: Dua puluh empat tikus putih jantan dewasa Strain Sprague Dawley dengan berat badan antara 200–300 gram dikelompokkan menjadi 4 grup (n=6). Grup kontrol negatif (CN) diberikan injeksi NaCl 0.9% 1cc secara intraperitoneal, sedangkan pada grup kontrol positif (CP) diberikan injeksi cisplatin 5 mg/kgBB pada akhir minggu ke-3. Dua grup lainnya masing-masing diberikan injeksi cisplatin 5 mg/kgBB dan vitamin E dengan dosis 50 mg/kgBB (P1) atau 200 mg/kgBB (P2). Aspirasi darah kardiak dilakukan di akhir minggu ke-7 dan diproses untuk analisa kadar testosteron. Hasil: Didapatkan kadar testosterone yang lebih rendah secara signifikan pada kelompok yang hanya diberikan injeksi cisplatin 5 mg/kgBB intraperitoneal dibandingkan dengan kelompok CN yang hanya diberikan NaCl 0.9% intraperitoneal (p=0.006), kelompok yang mendapatkan cisplatin 5 mg/KgBB dan vitamin E 50 mg/kgBB (p=0.003), dan kelompok yang mendapatkan cisplatin 5 mg/KgBB dan vitamin E 200 mg/kgBB lebih tinggi dibandingkan dengan kelompok yang mendapatkan cisplatin 5 mg/KgBB dan vitamin E 200 mg/kgBB. Simpulan: Paparan cisplatin menurunkan kadar testosteron pada tikus putih, dan pemberian vitamin E dapat memperbaiki pengaruh cisplatin terhadap kadar testosteron darah.

*Kata Kunci:* Cisplatin, kemoterapi, vitamin E, antioksidan, testosterone, testis.

Correspondence: Dian Kartika Rezia; c/o: Department of Urology, Faculty of Medicine/Universitas Airlangga, Soetomo General Hospital, Surabaya. Jl. Mayjen. Prof. Dr. Moestopo 6-8 Surabaya 60286. Phone: +62 31 5501318; Fax: +62 31 5024971. Mobile phone: 083857857353. Email: diankrezia@gmail.com.

# INTRODUCTION

Reports estimated about 14.1 million new cancer cases and 8.2 million cancer-related deaths worldwide in 2012. Of those figures, 57% of new

cases and 65% of deaths were reported in developing countries. The differences in these numbers between developed and developing countries are probably due to the different clinical features of patients, as well as diagnostics and therapeutics facilities. One

percent of new cancer cases in the United States were accounted for juvenile (0-14 years) and adolescence (15-19 years). Testicular cancer is marked on the fourth place for adolescence malignancy in the US.<sup>2</sup>

Cisplatin is one of the most potent and commonly used chemotherapy agent, especially in testicular, bladder, cervix, nasopharyngeal, and lung cancers. Despite its wide potency, cisplatin is also known to have numerous toxicity, including on testicular tissues. Studies have described negative effect of cisplatin on testicular tissue, both on its spermatogenic and steroidogenic functions. A study by Kilarkaje demonstrated a disturbance in the synthesis of testosterone in rats receiving 3 cycles of BEP with or without antioxidant administration. Betalance in the synthesis of testosterone in rats receiving 3 cycles of BEP with or without antioxidant administration.

Intracellular cisplatin causes an increase in reactive oxygen species (ROS) which will induce numerous chain reactions including apoptosis and cell necrosis. Physiologically, spermatozoa will produce a small amount of ROS for intracellular regulation, capacity, and acrosome reactions. However, if produced excessively, ROS in testicular tissue will cause damage to cellular elements and DNA fragmentation. Testicular tissue is highly sensitive of ROS and free radicals. Binding of free radical to a testicular component (spermatogonium, Sertoli cells, or Leydig cells) can induce lipid peroxidation and other chain reactions leading to apoptosis and cell necrosis." Previous studies have demonstrated a dysfunction of Levdig cells after exposure to cisplatin, both in short or long-term, though not all show marked decrease in testosterone levels. 13-16

Vitamin E is known to be a potent chain-breaking antioxidant, as well as lipid-soluble. Traber and Stevens stated that vitamin E indirectly prevents the formation of free radicals, or the early oxidation of fatty acids, and stopping the lipid peroxidation chain-reaction. Administration of vitamin E ( $\alpha$ -tocopherol) is expected to prevent lipid peroxidation reaction in cell membrane caused by ROS as a byproduct of cisplatin, thus an improved level of testosterone.

#### **OBJECTIVE**

This study is aimed to observe the difference in testosterone levels in white rats (Sprague Dawley strain) receiving combination of cisplatin and vitamin E compared to those receiving only cisplatin.

## **MATERIAL & METHODS**

In this laboratory experimental study we analyzed serum testosterone level after rats were given treatments. Twenty-four adult male rats (strain Sprague Dawley) weight 200-300 grams were assigned randomly into 4 experimental groups (n=6). Rats in the Negative Control (NC) group were given distilled water orally for 7 weeks and injected 1 ml of normal saline at intraperitoneally at the end of the 3<sup>rd</sup> week. Rats in the Positive Control (PC) group were given distilled water orally for 7 weeks, and were injected 1 ml of cisplatin (Kalbe Farma, Indonesia) 5 mg/kgBW intraperitoneally at the end of the 3<sup>rd</sup> week.

In therapy group 1 (P1) and therapy group 2 (P2) rats were respectively given 0.5 ml vitamin E 50 mg/kgBW and 200 mg/kgBW orally for 7 weeks, and were injected 1 ml of cisplatin 5 mg/kgBW intraperitoneally at the end of the 3<sup>rd</sup> week. The vitamin E doses were based on a previous study by Gevrek and Erdemir, which showed the same level of protection against oxidative stress on testis and sperm, without any significant side effects reported. At the end of the 7<sup>th</sup> week cardiac blood was aspirated and then tested for testosterone level using ELISA. This study has been ethically approved by the Veterinary Faculty of Universitas Airlangga, Surabaya.

## **RESULTS**

We studied 24 male adult rats aged 10-12 weeks, which characteristics are shown in table 1.

Shapiro-Wilk analysis showed a normal data for testosterone levels in each group, and with a homogenous variance (p>0.05). Based on those results, we analyzed the data further using one-way ANOVA, as shown in table 2. Our data showed a significant difference in testosterone levels between groups (p<0.05), thus we used the LSD post hoc test to compare each groups.

We found a significantly lower testosterone levels (p<0.05) in the positive control group (CP) compared to the negative control group (CN). Testosterone levels were also significantly lower in the CP group compared to those in the P1 and P2 groups (p=0.003 and p=0.001, respectively). Although statistically insignificant, we also found that the testosterone levels in P2 group were lower than in P1 (p=0.702).

**Table 1.** Basic sample characteristics.

Weight	$Mean \pm SD$	p
Pre Treatment (gram)		
CN	$248.33 \pm 18.34$	$0.57^{a}$
CP	$252.50 \pm 23.18$	
P1	$249.17 \pm 30.07$	
P2	$263.33 \pm 27.33$	
Testis Weight (gram)		
CN	$1.40 \pm 0.18$	$0.74^{a}$
CP	$1.33 \pm 0.21$	
P1	$1.31 \pm 0.22$	
P2	$1.25 \pm 0.23$	

CN: Control Negative

CP: Control Positive (Cisplatin 5 mg/kgBB)

P1 : Cisplatin 5 mg/kgBB + Vitamin E 50 mg/kgBB

P2: Cisplatin 5 mg/kgBB + Vitamin E 200 mg/kgBB

a : Kruskal-Wallis Test

\* : statistically significant if p<0.05

**Table 2.** Comparison of testosterone levels among groups.

Group	n	$Mean \pm SD$	р
CN	6	$6.353 \pm 0.229$	0.004*
CP	6	$5.899 \pm 0.070$	
P1	6	$6.406 \pm 0.259$	
P2	6	$6.461 \pm 0.370$	

<sup>\*</sup> p<0.05 = statistically significant

**Table 3.** LSD post hoc analysis of testosterone levels between groups.

Comparison between groups	Mean Difference	CI 95%		n voluo
		Lower Bound	Upper Bound	p value
CN vs CP	0.454	0.147	0.762	0.006*
CN vs P1	-0.052	-0.360	0.255	0.726
CN vs P2	-0.109	-0.417	0.198	0.466
CP vs P1	-0.507	-0.814	<b>-</b> 0.199	0.003*
CP vs P2	-0.564	-0.872	-0.257	0.001*
P1 vs P2	-0.057	-0.364	0.250	0.702

<sup>\*</sup> p<0.05 = statistically significant

## **DISCUSSION**

Cisplatin, or Cisdiammine dichlorido platinum (II), is considered one of the most widely used chemotherapy agents. Its potency has been known for testicular cancer, and other types of cancers. Unfortunately, along with its benefits as anti-tumor agent, this drug also causes several toxicities in both short and long-term use on various

organs and is known to be dose-related. The most commonly occurred toxicity is on the kidneys, which is caused by severe degeneration of the glomeruli and distal tubules. Other studies also reported other long-term toxicities such as ototoxicity, neurotoxocity, and myelosuppression. 3,5,6

In testicular tissues, cisplatin have also been reported to cause both spermatogenic and steroidogenic dysfunction. 8-10 Oxidative stress caused by an

increase in concentration of ROS and RNS as the product of cisplatin on testicular tissue is marked by elevated lipid peroxidation and a reduction of antioxidant system, leading to changes in the tissue histologically. 22,23 Animal subjects exposed to cisplatin are reported to suffer from testicular damage indicated by germinal cells apoptosis, Levdig cells dysfunction, and a decrease in steroidogenic function. Cisplatin is known to inhibit the synthesis of nucleic acid in germinal cells, resulting in disruption of spermatogenesis, while also causing LH receptors dysfunction and damaging Leydig cells viability - which results in the decreased testosterone production - both to be blamed for infertility at a later point. A study by Ciftci detailed the spermatogenic dysfunctions caused by cisplatin as azoospermia, dysmorphology of the sperm, decreased sperm motility, and disrupted spermatogenesis. <sup>13,22,24,25</sup>

In the current study, single intraperitoneal administration of cisplatin 5 mg/kgBB caused a significant decrease of testosterone levels compared to control group (p<0.05). This result is similar with a study by Aydiner et al., which showed a marked decrease in testosterone level, followed by an increase in Leutenizing Hormone (LH) level on the 3<sup>rd</sup> and 21<sup>st</sup> day after administration of cisplatin. Ultrastructural damages of the Leydig cells, such as dilatation of the endoplasmic reticulum (ER) and increase in lipid and lipofusion inclusions, as well as mitochondrial structural disorganizations were observed under electron microscopy on the 3<sup>rd</sup> until the 21<sup>st</sup> day after cisplatin injection.<sup>26</sup>

As has been understood, the chain reaction of testosterone biosynthesis takes place in the mitochondria, as well as the smooth endoplasmic reticulum. A study by Maines (1990) demonstrated a significantly decreased level of P450scc as well as its activity in rats treated with cisplatin. <sup>27</sup> Garcia (2012) demonstrated cisplatin exposure on cultured testicular interstitial cells in several modified microenvironment. It was also found that cisplatin causes an acute significant decrease in P450scc enzyme which was followed by a decrease in testosterone synthesis. The decrease, as explained in the study, was unlikely to be caused by the downregulation of the enzyme which has a slow turnover. Moreover, Cisplatin did not seem to have any effect on the P450scc mRNA levels. Thus, it was concluded that the inhibition of the steroid synthesis is caused by a ROS-mediated mechanism.<sup>28</sup>

Marked improvements were found in testosterone levels of rats receiving vitamin E. In this study we found a significantly higher level of testosterone with the administration of vitamin E 50 mg/kgBW (P1) compared to those only injected with cisplatin 5 mg/kgBW (CP)  $(6.406 \pm 0.259 \text{ vs. } 5.899 \pm 0.070, p=0.003)$ . The same result was also seen in the group receiving higher dose of vitamin E (P2) compared to CP  $(6.461 \pm 0.370 \text{ vs. } 5.899 \pm 0.070, p=0.001)$ . However, the higher dose of vitamin E administered did not seem to give significantly better improvement of testosterone levels (p=0.702).

Platinum molecules from cisplatin will bind to the intracellular DNA and produce reactive oxygen species (ROS), which will cause several chain reactions, including lipid peroxidation.<sup>3,6</sup> Vitamin E (α tocopherol) is potent lipid-soluble antioxidant and works as a potent chain-breaking scavenger for peroxyl radicals and inhibits the propagation of free radicals in cell membrane and plasma lipoprotein. The OH atom from α tocopherol is phenolic, and binds to a peroxyl radical to form hydroperoxide. Tocopheroxyl radical formed in this reaction is relatively stable, non-radical, and inactive. 18,29 Traber and Stevens stated that vitamin E indirectly prevents the formation of free radicals, or the early oxidation of fatty acids, and stopping the lipid peroxidation chain-reaction.<sup>17</sup> However, in a high concentration α tocopherol may also induce the formation of lipid hydroperoxide, which can bind to a lipid free radical and eventually cause membrane damage as well. 18,30

A study by Umeda in 1978 demonstrated an activation of smooth ER development in rat's Leydig cells with administration of vitamin E. Furthermore, Umeda reported in his study in 1982 that there was a significant increase of testosterone levels in both plasma and testicular tissue after administration of vitamin E in a long-term. This might explain the basic mechanism of testosterone improvement in rats treated with cisplatin. Vitamin E works as an antioxidant which prevents lipid peroxidation and damage to Leydig cells and its components, thus ensuring the biosynthesis of testosterone. Moreover, vitamin E also induces the development of smooth ER which plays an important role in testosterone biosynthesis.

Despite the obvious positive effect of vitamin E on testosterone levels in mice, this study does not fully represent the actual condition of patients with malignancy receiving cisplatin therapy.

Firstly, because it was designed as a post-test only study, we cannot compare the testosterone levels before and after the administration of vitamin E, with and without cisplatin, and vice versa. Secondly, we only administered a single dose of cisplatin, instead of multiple dose with interval as dictated on chemotherapy protocols.

## **CONCLUSION**

Administration of vitamin E helps recover the negative effect caused by cisplatin on testosterone levels.

#### REFERENCES

- 1. Torre LA, Bray F, Siegel RL. Global cancer statistics 2012. CA Cancer J Clin. 2015; 65: 87-108.
- 2. Ward E, DeSantis C, Robbins A. Childhood and adolescent cancer statistics. CA Cancer J Clin. 2014; 64: 83-103.
- 3. Riddell Imogen A, Lippard Stephen J. Cisplatin and oxaliplatin: Our current understanding of their actions. In Sigel Astrid, Sigel Helmut, Freisinger Eva, Sigel Roland KO. Metallo-Drugs: Development and Action of AnticancerAgents. Berlin: de Gruyter GmbH; 2018. p. 1-42.
- Mieog JSD, van der Hage JA, van de Velde CJH. Neoadjuvant chemotherapy for operable breast cancer. British Journal of Surgery. 2007; 94(10): 1189-200.
- 5. Jafri SH, Mills G. Neoadjuvant chemotherapy in lung cancer. Therapy. 2011; 8(1): 23-31.
- 6. Barabas K, Milner R, Lurie D. Cisplatin: a review of toxicities and therapeutic applications. Vet. Comp. Oncol. 2008; 6: 1-18.
- 7. Hoff PM, Saad ED, Costa F. Literature review and practical aspects on the management of oxaliplatin-associated toxicity. Clin Colorectal Cancer. 2012; 11(2): 93-100.
- 8. Howell SJ, Shalet SM. Testicular function following chemotherapy. Hum Reprod Updat. 2001; 7: 363-9.
- 9. Colpi GM, Contalbi GF, Nerva F. Testicular function following chemo-radiotherapy. Eur J Obstet Gynecol Reprod Biol. 2004; 113(Suppl. 1): S2-6.
- 10. Kilarkaje N. Effects of combined treatment of α tocopherol, l ascorbic acid, selenium and zinc on bleomycin, etoposide and cisplatin induced alterations in testosterone synthesis pathway in rats. Cancer Chemother Pharmacol; 2014.DOI 10.1007/s00280-014-2592-8
- 11. Agarwal A, Makker K, Sharma R. Clinical relevance of oxidative stress in male factor infertility: an update. Am J Hum Reprod Immunol. 2008; 59: 2-11.
- 12. Ishikawa T, Fujioka H, Fujisawa M. Clinical and hormonal findings in testicular maturation arrest.

- BJU Int. 2004; 94: 1314-6.
- 13. Soni KK, Kim HK, Choi BR, Karna KK, You JH, Cha JS, et al. Dose-dependent effects of cisplatin on the severity of testicular injury in sprague dawley rats: reactive oxygen species and endoplasmic reticulum stress. Drug Des Devel Ther. 2016; 10: 3959-68.
- Strumberg D, Brugge S, Korn MW. Evaluation of long-term toxicity in patients after cisplatin-based chemotherapy for non-seminomatous testicular cancer.
- 15. Sprauten M, Brydey M, Haugnes HS. Longitudinal serum testosterone, luteinizing hormone, and follicle-stimulating hormone levels in a population-based sample of long-term testicular cancer survivors. J Clin Oncol. 32: 571-8.
- 16. Bandak M. Longitudinal changes in serum levels of testosterone and luteinizing hormone in testicular cancer patients after orchiectomy alone or bleomycin, etoposide, and cisplatin. Eur Urol Focus; 2016. http://dx.doi.org/10.1016/j.euf.2016.11.018
- 17. Traber MG, Stevens JF. Vitamins C and E: beneficial effects from a mechanistic perspective. Free Radical Biology & Medicine. 2011; 51: 1000-13.
- 18. Yamauchi R, Miyake N, Kato K, Ueno Y. Reaction of c!-tocopherol with alkyl and alkylperoxyl radicals of methyl linoleate. Lipids. 1993; 2(8): 201-6.
- 19. Gevrek F, Erdemir F. Investigation of the effects of curcumin, vitamin E and their combination in cisplatin-induced testicular apoptosis using immunohistochemical technique. Türk Üroloji Dergisi/Turkish J Urol. 2018; 44: 16-23.
- Cepeda V, Fuertes M, Castilla J, Alonso C, Quevedo C, Pere, J. Biochemical mechanisms of cisplatin cytotoxicity. Anticancer: Agents Med Chem. 2007; 7: 3-18.
- 21. Pisano C, Pratesi G, Laccabue D. Paclitaxel and cisplatin-induced neurotoxicity: a protective role of acetyl-1-carnitine. Clin Cancer Res. 2003; 9:5756-67.
- 22. Ciftci Osman. Comparison of reproductive toxicity caused by cisplatin and novel platinum-nheterocyclic carbene complex in male rats. Basic Clin Pharmacol Toxicol; 2011. p. 328-33.
- 23. Martins NM, Santos NA, Curti C, Bianchi ML, Santos AC. Cisplatin induces mitochondrial oxidative stress with resultant energetic metabolism impairment, membrane rigidification and apoptosis in rat liver. J Appl Toxicol. 2008; 28(3): 337-44.
- 24. Vawda AI. Effect of testosterone on cisplatin-induced testicular damage. Arch Androl. 1994; 32(1): 53-57.
- Azouri H, Bidartt JM, Bohuon C. In vivo toxicity of cisplatin and carboplatin on the leydig cell function and effect of the human choriogonadotropin. Biochem Pharmacol. 1989; 38: 567-71.
- 26. Aydiner A, Aytekin Y, Topuz E. Effects of cisplatin on testicular tissue and the leydig cell-pituitary axis. Oncology. 1997; 54: 74-78.
- 27. Maines MD, Sluss PM, Iscan M. Cis-platinummediated decrease in serum testosterone is associated

- with depression of luteinizing hormone receptors and cytochrome P-450scc in rat testis. Endocrinology. 1990; 126: 2398-406.
- 28. Garcia MMS, Acquier A, Suarez G, Gomez NV. Cisplatin inhibits testosterone synthesis by a mechanism that includes the action of reactive species oxygen (ROS) at the level of P450scc. Chemico-Biological Interactions. 2012; 199: 185-91.
- 29. Terao J, Matsushita S. The peroxidizing effect of  $\alpha$ -tocopherol on autoxidation of methyl linoleate in bulk phase. Lipids. 1986; 21:255-60.
- 30. Akazawa N, Mikami S, Kimura S. Effects of vitamin E deficiency on the hormone secretion of the pituitary-gonadal axis of the rat. Tohoku J Exp Med. 1987; 152(3): 221-9.
- 31. Umeda F. Functional and morphological studies on pituitary-gonadal, and adrenal axes in vitamin E deficient and supplemented male rats. Fukuoka Acta Med. 1978; 69: 327-37.
- 32. Umeda F, Kato K, Muta K, Ibayashi H. Effect of vitamin E on function of pituitary-gonadal axis in male rats and human subjects. Endocrinol Jpn. Jun 1982; 29(3): 287-92.